REVIEW- PROTEIN ISOLATES: PRODUCTION, FUNCTIONAL PROPERTIES AND APPLICATION

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ABSTRACT:
Protein isolates are refined form of protein containing the greater amount of protein with greater digestibility. Nowadays it’s the major source of cheap proteins especially for athletes, bodybuilders, vegetarians, and has gained wide application in various beverages and dairy industries, and infant foods due to its different functional properties. It is widely produced from deoiled cake of legumes such as peanuts, cowpeas, soybeans, etc and animal sources like fish and milk by a combination of isoelectric and alkaline precipitation using H₂SO₄ and NaOH, followed by centrifugation to obtain the pure isolates which can further be dried to powdered form of about 90% protein. The problem of antinutritional factors in legumes can be successfully reduced by these processing methods.

KEY WORD: Protein Isolates, isoelectric precipitation, Ultrafiltration antinutritional factor.

INTRODUCTION:
Proteins are nitrogen-containing compounds made of up amino acids unit. They are the major structural component of muscles and other tissues in the body. They are also component of hormones, enzymes and haemoglobin (Jay et al.2004). Proteins composed of twenty different amino acids linked together by a peptide bond and the resulting chain is called polypeptide. They have similar basic structure but differ in their side chains. This difference in side chains gives the proteins their specificity and functionality. These amino acids are classified as essential (those that cannot be synthesis by body but rather taken as...
supplement from diet), and nonessential (that can be synthesis by the body) (Clark 2003).

Demand for the relatively cheap sources of protein that can be incorporated to value-added food products is increasing worldwide, and numerous researches are still going on various sources of plant proteins that may help in improving the nutritional value of food products at low cost (Gurpreet et al. 2006).

Proteins are available in different varieties of dietary sources including animals, plants origin, and from highly marketed spot supplement industry. Typically, all dietary animal proteins (e.g. eggs, milk, meat, fish and poultry) are considered complete protein because they contain all essential amino acids. Proteins from vegetable sources (such as legumes, nuts and soy) are incomplete proteins since they are lacking one or two essential amino acids.

Assessing the quality of protein is important when considering the nutritional benefit that it can provide. Protein quality of food is the ability of the food to meet the nutritional requirement of an individual species. It is indicated by how well the protein is digested, absorbed, and utilised for the growth and sustenance of the body (Wardlaw et al. 1996). The quality of protein can be determined by assessing its essential amino acid profile. Numerous methods to determine the quality of proteins include :

1) Protein efficiency ratio (PER) which is achieved by measuring the animal growth after feeding with the test protein. The weight gained in grams per gram of protein consumed is calculated and then compared with the standard value of 2.7 (standard value of casein protein). If the value is greater than 2.7, the food is regarded as excellent source of protein. However, the result mostly provides a measure in rat but does not provide a strong correlation to human growth needs.

2) Biological value is calculated by dividing the nitrogen used for tissue formation and the nitrogen absorbed from the food and then multiplied by 100 and express as percentage of nitrogen utilised. This gives an idea of how efficient the body utilises the protein consumed in the diet. Higher value indicates that food supplies high essential amino acids.

3) Net protein utilisation (NPU) is almost identical to BV except that direct measure of retention of absorbed nitrogen is involved. However, the BV is calculated from nitrogen absorbed while NPU is calculated from nitrogen ingested (Jay et al. 2004).

4) Protein digestibility Corrected Amino acids Score, this, according to joint (FAO/WHO, 1990) could be determined by expressing the content of the first limiting essential amino acid of the test protein as a percentage of the content of the same amino acid content in a reference pattern of essential amino acid. The reference values used were based on essential amino acid requirement of preschool-age children, which was corrected for true faecal digestibility of the test protein. The value obtain was known as protein digestibility corrected amino acid score (PDCAAS).
Protein isolates
Isolates are the most refined form of protein products containing the greatest concentration of protein but unlike flour and concentrates contains no dietary fibre. Isolates originated from United States around 1950s (Jay and Michael 2004). They are very digestible and easily incorporated into different food products. Protein isolates are nowadays believed to have played a major role in the development of new class of formulated foods. It is high concentration of protein with the advantage of colour, flavour and functional properties make it an ideal raw ingredient for use in beverages, infant foods and children milk food, textured protein products and certain types of specialty foods (Olaofe 1998). Protein isolates have been developed from a variety of legumes among which are soy bean, peanut, canola, cashew nut, almonds, sesame, pinto and navy beans (Seyam et al. 1983).

Extraction Methods
Isoelectric precipitation: - The isoelectric point (pI) of a protein is the pH where the net charge on the protein is zero. Proteins tend to aggregate and precipitate at their pI because there is no electrostatic repulsion keeping them apart. Proteins have different isoelectric point because of their different amino acid sequence, and therefore, they can be separated by adjusting the pH of a solution. When the pH is adjusted to the isoelectric point of a particular protein it precipitates leaving the other protein in the solution (McClements 2013). A more recent study on isoelectric precipitation of soya protein confirmed that both extraction condition and the type of acid used for isoelectric precipitation had little effect. However, it was found that an isoelectric point of around pH 4.2 leads to high proportion of protein precipitated out (Smith et al. 1939).

Alkaline extraction: It was found that alkaline reagents were more effective in extraction of protein from food legumes. However, several changes such as destruction of lysine, formation of lysine-alanine, racemisation may occur during alkali extraction and reduce the protein quality (Tobin et al. 1978). Hence, a better technology to produce protein isolate and concentrate with maximum yield and minimum or no anti-nutritional factors need to be developed.

Ultrafiltration method: - This is a potential membrane processing for extraction of protein, a solution of protein is placed in a cell containing a semi-permeable membrane, and pressure is applied. Small molecules pass through the membrane, whereas the larger ones remain in the solution. The semi-permeable membranes with cut-off points of between 500 and 300,000 are mostly available (McClements 2013). The extraction of soy protein isolate by this method was first reported over twenty years ago. The Ultrafiltration system provides a commercially feasible alternative to the existing isoelectric precipitation method of soya beans production. Subsequently, many manufacturers were
believed to have implemented the ultrafiltration process for soya isolate. However, very few details are available due to commercial confidentiality (Smith et al. 1939). Ultrafiltration can also be used to concentrate protein solution and can be used in laboratory and commercial scale (McClements 2013).

Figure 1: Systematic flow sheet for legume protein isolates recovery (Qayyum et al. 2012)

Protein isolate from different plant and animal sources
Proteins that are utilised in food processing are of various origins, and can roughly be classified into animal proteins (gelatins), vegetable proteins (e.g. peanut protein, soy protein, wheat proteins, Almond protein, canola meal protein etc.), and animal derived protein (e.g. milk proteins). However, many vegetable proteins require processing to provide food material having acceptable functional properties such as emulsification, oil and water absorption, texture modifications, colour control and whipping properties, which are primarily attributed to the protein characteristics. Many plants have attracted a deal of interest as a source of low-cost protein to supplement human diet, this include among others soybeans and peanut (Ogunwolu et al. 2009).

Whey protein isolates (WPI)
Whey is the liquid by-product of cheese which can further be processed into a spray dried products like instance whey protein concentrates (WPC), whey protein isolate (WPI) or whey protein hydrolysate (WPH) (Brucic et al. 2009). By definition from the code of Federal Regulation, is the liquid substances obtained by separating the coagulum from milk or cream in the cheese making. It represent about 20
percent of the milk proteins, which remain in the serum phase during processing of cheese (Kimberlee 2012). During the production of whey protein isolates, significant amount of fat and lactose get removed, as a result of which individual with lactose-intolerant can safely consume these products\(^1\). Whey proteins are widely used as ingredients in different foods (dairy, meat and bakery products) due to their unique functional and nutritional properties (Brucic et al. 2009). One specific aspect of whey protein that may be challenging for some formulations is sensitivity to heat. Consequently, the Dairy Research Institute, established under the leadership of America’s dairy farmers through the dairy check off program, has supported a variety of research that aims to improve whey protein’s performance in higher heat processing (Kimberlee 2012). Beta-lactoglobulin and alpha-lactalbumin are the major whey protein responsible for heat stability characteristics of ingredients such as WPC and WPI (Vardhanabhuti and Foegeding 2008). The relationship between sugar addition and heat stability of whey protein isolate have been studied by many researchers. Sucrose addition was found to increase gelation temperature and gel strength of WPI and bovine serum albumin (Rich and Foegeding 2000). Adding glycerol improved heat stability of WPI and decreases turbidity and protein gelation (Kulmyrzaev et al. 2000). Thermal denaturation temperature of WPI was also found to increased by addition of sorbitol more effectively than glycerol (Paredes-Lopez 2006). Many food application contains sugars or sugar alcohols, and their presence can help to improve the heat stability of whey protein ingredients by preventing the formation of large aggregates and providing better clarity in application such as beverages (Kimberlee 2012).

**Fish Protein isolates (FPI)**

To solve the problem of utilisation of unconventional raw material (dark muscle fish, fatty fish) and also fish by-products (fish trims, fish frames etc.) a process was developed to economically develop a functional protein isolates from these kinds of raw materials. This technology uses the pH dependant’s solubility properties of fish muscle proteins for their separation and recovery from other components of muscle that are not desirable in a final product. Fish protein isolate is a protein concentrate which is prepared from fish muscle without retaining the original shape of the muscle. It is not generally consumed directly, but used as raw material for production of other value added products. Fish protein isolate does not retain the original shape of muscle, and is normally utilized as ingredient for the production of value added products. It is still a good source of protein for the production of ready to eat fish products. The overall processes involved are simple. The proteins of the muscle tissue are first solubilised. The solubilisation can be accomplished by addition water with alkali added to approximately pH 10.5 or higher, or with acid added to about pH 3.5 or lower. It is usually necessary to
choose the pH at which the consistency of the solution decreases to a value that allows the removal of undesirable material. The mixture is then centrifuged, and due to density differences the oil rises to the top and can then be removed. Other insoluble impurities such as bone or skin are also sedimented at this stage. The muscle protein are then precipitated and collected by a process such as centrifugation (Shaviklo 2006).

**Peanut protein isolates (PPIs)**

Peanut (*Arachis hypogaeae L.*) is an annual herbaceous plant belonging to the suborder *papillonacea* of the order *Leguminosea*. It contains 26-29% protein with good nutritional quality. Peanut proteins are used for their functional properties (emulsification, forming) or for their nutritional properties in different food products. They are also used for human nutrition in developing countries to supplement cereals, beverages and skim milk. Peanut protein isolate can be prepared from the defatted peanut cake or powdered by macerating with high salt phosphate buffer (20mM Na$_2$HPO$_4$, 2mM KH$_2$PO$_4$, 5.4 mM KCl, 1M NaCl, pH 7.4), centrifuging and then supplementing the supernatant with (NH$_4$)$_2$SO$_4$ to 90% saturation. After centrifuging the pellet can be dialyse against distilled water overnight at 4oC and freeze-dried (Mouecoucou 2004).

**Soy protein isolates (SPIs)**

Soy protein isolate is a common isolate. It has high protein content of about 90%. It is made of defatted soy meal by removing most of the fat and carbohydrates (Seyam 1983). A soybean is crushed into oil and defatted meal. The meal is usually used as animal feed, while smaller amount is further processed into food ingredients including soy flour, protein concentrate, protein isolates and textured protein (Kinsella 1976). Soy protein isolate is usually combined with other food ingredients such as vitamins, minerals and flavour in preparation of soy protein shake powder (Seyam 1983). Advance in food technology resulted in the development of a variety of soy product such as concentrates, isolate and extruded-expanded products, this consequently leads to increased utilisation by technically developed regions of the world (Young 1979). The production of soy protein isolate involve solubilising the protein and carbohydrate at neutral or alkaline pH and the recovery of the solubilised protein, separation and optionally washing and neutralization before drying (Moure et al. 2006). Three steps involved in the processing of soy protein isolates (SPI) are (1)The soy flakes are slurried with water under alkaline conditions (pH 6.8-10 at 27-66oC using sodium hydroxide and other alkaline substances approved for food used) so that the protein and the oligosaccharides can dissolve into the solution. The protein solution is then separated from the insoluble residue by centrifugation, (2) the supernatant containing the protein and sugars is then acidified to isoelectric pH 4.5 ( where the solubility of proteins
is minimal), using hydrochloric acid (HCl). This leads to the precipitation of protein as curd, (3) the solubility of the precipitated protein is restored by neutralizing to alkaline pH of 6.5-7.0 after rediluting with fresh water or spray dried in its acidic form and packed in multilayer paper bags (Lusas et al. 1995, and Anon 2008).

**Canola protein isolates (CPIs)**

According to USDA 2010, Canola meal has been the second largest feed meal after soybean meal. It has a good amino acid profile with a well balance amino acid composition although it has found only marginal used in the food industry, due to the presence of anti-nutritional factors. The vast majority of canola protein isolates are prepared by alkaline extraction method followd by isoelectric precipitation. Although, this extraction method generates high yield of nitrogen, but the isolate produced by this method have been found to have poor solubility and digestivity, this is most probably due to the nature of proteins constituent of canola meal which consist of alkaline-soluble fraction that can be easily denatured during the extraction process. CPI thus possesses generally an unacceptable food-functional properties including poor water holding, gelling and oil-binding, foaming and emulsification properties. Many studies have been carried out to modify the properties of this isolate e.g. by succinylation, acylation and enzymatic hydrolyses. However, enzymatic hydrolyses has been prepared because it’s less effect than chemical method (Alashi 2011).

**Chick pea protein Isolate**

Chick pea protein is the world’s third largest pulse crop in term of area, grown mostly in West Asia and Mediterranean region. It is one of the major vegetable proteins. Many functional properties of this protein isolate has been studied whereas, information on gelation properties of chick pea protein isolate was scares. The chick pea protein isolate dispersed with sodium and calcium salts showed different rheological behaviour at different ionic strength and pH. Increasing the ionic strength of dispersion could strengthen the gelation properties of CPI under acidic conditions, however reduced the elastic parameters of CPI at pH of 7.0 (Zhang 2007).

**Cashew nut protein isolate**

Cashew has considerable economic importance because its components have numerous economic uses. The kernel has high food value with about 40-57% oil and 21% protein content. A cashew kernel meal contains about 42% crude protein, a low crude fibre and 0.5% and 0.2% calcium and phosphorous, respectively, which is comparable to that of peanut composition, which has been used for peanut protein isolate and concentrate. Protein isolates and concentrates can be obtained from defatted cashew nut powder by both alkaline extraction-iseoelectric precipitation (IP) and alkaline extraction-
methanol precipitation (MP). Cashew nut protein isolates has water and oil absorption capacities of 2.20ml/g and 4.42ml/g respectively, emulsifying stability index (447%), foam capacity and stability (45% and 55%, respectively), and low gelation capacity of (13.5%) (Ogunwolu 2009).

**Functional Properties of Protein Isolate**

**Bulk density:** - Bulk density is an important parameter that determines the packaging requirement of a product. It signifies the behaviour of a product in dry mixes and can varied with fineness of particles (Butt and Batool 2010). The bulk density of protein isolate of sources can be determined by taking 10 grams of protein isolate into 100ml graduated cylinder and taping several time on a laboratory bench until the isolate settled and then taking the values, which can be express as g/cm (Olaofe 1998, and Okaka and Potter 1977).

**Protein solubility:** - Protein solubility is a useful indicator for the performance of protein isolates incorporated in the food system and also the extent of protein denaturation because of heat and chemical treatment at different pH (Horax 2004). To determine the protein solubility, the protein isolates (250mg) should be homogenized in 20ml of 0.1M NaCl at pH of 7.0 for 1h fallowed by centrifugation at 10,000×g for 30min. Nitrogen content should then be determined in the soluble fraction and then solubility can be expressed as the percentage of total nitrogen of the original sample to that of soluble fraction (Morr 1985).

**Foaming Capacity and stability:** - The foaming properties are used as an index of the whipping characteristics of the protein isolate (Mwasaru 1999). The capacity and stability of foams can be determined by dispersing 50ml of 3%(w/v) dispersions of protein isolate sample in distilled water and immediately transfer into a graduated cylinder, the volume should then be recorded before and after whipping. The foaming capacity can be expressed as the percentage volume induced by whipping. The change in volume of foam after 60 min of standing at room temperature is recorded as foam stability (Lin 1974).

**Water/oil absorption:** Proteins has both hydrophilic and hydrophobic properties therefore, can interact with water and oil in foods (Butt and Batool 2010). The lower water absorption capacity can be due to less availability of polar amino acids (Kuntz 1971), and the low fat absorption may be due to the presence of large proportion of hydrophilic and polar amino acids on the surface of the protein molecules (Sathe et al.1982).

Water absorption can be determine by mixing approximately 3g of protein isolate with 25ml distilled water in a pre-weighed centrifuge tubes, the tubes are then stirred and centrifuge for 25 min at 3000×g for 30 min interval. The supernatant should be after by 25 min drainage at 50°C, then re-whining the
protein isolate. Water absorption is express as the number of grams of water absorbed per gram of sample (Sosulski et al. 1976). High water absorption of protein isolates help to reduce moisture loss in package bakery products (Lin 1974).

Oil absorption can also be estimated by mixing 0.5g of protein isolates with 6ml corn oil in pre-weighed centrifuge tubes. The tubes are then stirred for like one minutes to get complete dispersion of the sample in oil. After 30 min holding time, the samples are centrifuge at 3000×g for 25 min. The supernatant should be with pipette and invert the tubes for 25 min to drain the oil before re-weighing. The oil absorption is express as gram of oil absorbed per gram of protein isolate (Sosulski 1976). The high oil absorption is essential in the formulation of food systems like sausages, cakes batters, and mayonnaise and salad dressing (Butt and Batool 2010).

**Emulsifying activity and stability:** - Proteins are surface active agents that can form and stabilize the emulsion by creating electrostatic repulsion on oil droplet surface (Makri et al. 2005). To determine the emulsifying activity, Protein isolate (3.5g) was homogenised for 30 seconds in 50ml water using homogenizer at approximately 10,000rpm. Corn oil (25ml) was added the mixture was homogenised for 30 sec. The emulsion is divided into two equal volume aliquots and centrifuged at 1100×g for 5 min, the aliquots is heated for 15 min at 85°C. The ratio of the height of emulsion to the high of liquid layer was noted to calculate emulsion activity. The emulsion stability remains after heating (Naczk et al. 1985).

**Least gelation concentration:** - This is a qualitative parameter that expresses the minimum protein concentration at which the gel does not slide along the test tube walls in inverted position (Young and Scrimshaw 2006). The lower gelation concentration the better is the gelling ability of proteins, because protein gels are aggregates of denatured molecules (Akintayo 1999).

**Antinutritional factor in protein isolate**

Several studies have reported the preparation and functional properties of protein isolates and concentrate (Sathe and Salunkhe 1981). Legumes contain some anti-nutritional factors such as lectins, saponin, haemagglutinin, protease inhibitor, oxalate, goitrogen, phytates, trypsin inhibitor and tannin (Amarowicz and Pegg 2008). These compounds reduce protein digestibility and availability. Some anti-nutritional in legumes have been reported to have health benefits. Tannin, a polyphenolic compound is reported to possess antioxidative property (Apata and Ologhobo 1997). Very extensive work has been carried out on optimising the conditions for trypsin inhibitor destruction during processing of soya protein isolate (Smith and Circle 1977). Studies had also shown that trypsin and chymotrypsin inhibitory activities were reduced to 75.3% and 84.8% in protein concentrates, 98.0% and 85.4% in
protein isolates of Great Northern beans respectively. However, these protein fractions did not contain oligosaccharides of the raffinose family (such as raffinose, stachyose, and verbascose) and hemagglutinating activities. Therefore, most of the antinutritional factors can possibly be eliminated by preparation of protein isolates and concentrates (Sathe and Salunkhe 1981). Specific studies were carried out on isoelectric process for protein isolate, where trypsin inhibitors levels were found to be dependent on the degree of heat treatment to which the raw materials had been subjected and on the pH used for isoelectric precipitation. Trypsin inhibitor levels in isolate produced by ultrafiltration process have also been studied and found to be fairly high; this might possibly be due to complexing of the relatively low weight trypsin inhibiting protein fractions with higher molecular weight proteins (Honig and Wolf 1987).

**Application of protein isolate in food systems.**

Protein isolates are the acceptable ingredients for dairy application due to their fine particle size and dispersibility. Emulsification, emulsion stability, colour and flavour are critical in dairy application. Isolates (especially soy proteins) are being used to fortify all type of pasta products such as macaroni, spaghetti, to improve the nutritional value etc (Sipos 2013).

Protein isolates are important sources of protein with high lysine content. Isolates from different legumes varied slightly in physiochemical and thermal properties. They are used as proteinaceous ingredients in many food products such as salad dressing, meat products and dessert. Whey proteins are mainly used in beverage applications, due to their health benefits (Kudre 2013).

The problem of poor flavour, mouth feel, texture, dryness and flavour associated with the use of soy flour and soy concentrate above 10% has been resolved by using soy isolate in meat loaves, sausage-type products for their emulsion-stabilizing effect, gelation, and moisture retention and improved effects on texture (Kinsella 1976). Soy protein is regarded as textured protein products use both in meat and vegetarian meat analog industry and thus, has good water holding capability (Riaz 2006). It is often used as meat extenders in comminuted meat products such as patties, fillings, meat sauces, meat balls, etc (Berk 1992).

Many soy protein isolates have been developed for providing different functional or physical properties to meet the requirement of various food systems. Soy protein isolates form firm, hard, resilient gels, unlike soy flour and concentrates that form soft and fragile gels (Riaz 2006). The firm-forming ability of soy protein isolates is important in meat products. When heat and pressure are applied the protein films fused together to form a firm, continuous, textured mass that can be sliced and used as meat substitutes. They also vary in their ability to form gels. Some are designed to form gel while others will not form gel at 14% solid content. Protein isolates are utilised in meat and baking application due to their water and fat absorption properties, the value for these properties for protein isolates ranges from 150 to 400% (Sipos 2013).
Neutralised protein isolates are often highly dispersible and form gel under appropriate aqueous conditions. They have both emulsifying and emulsion stabilising properties, and are excellent fat and water binders. For this reason, they are widely used in processed meat products, as well as both coarse and fine emulsions (e.g., patties, loaves and sausages) (Sipos 2013).

Different protein products such as whey protein, soy protein isolates, wheat gluten, rice bran protein, peanut protein, and cottonseed proteins where investigated for film development (Rhim 1998).

**CONCLUSION:**

Protein isolates shows a lot of potential to combat the problem of malnutrition. The underutilised plant and animal sources can be exploited in order to extract the proteins and make them available for used as food supplements. Research is needed compared the benefit of different protein isolates and their benefit as food supplements.

**ACKNOWLEDGEMENTS:**

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**REFERENCES:**


**Table 1: Protein content of some protein isolates**

<table>
<thead>
<tr>
<th>S/N</th>
<th>Protein Isolate</th>
<th>% Protein</th>
<th>Method</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Whey</td>
<td>&gt;90</td>
<td>Ion-exchange chromatography and membrane filtration</td>
<td>(Foegeding <em>et al</em>., 2011)</td>
</tr>
<tr>
<td>2</td>
<td>Chickpea flour</td>
<td>84.8-87.8</td>
<td>Isoelectric precipitation</td>
<td>(Paredes-Lopez <em>et al.</em>, 2006)</td>
</tr>
<tr>
<td>3</td>
<td>Lentil</td>
<td>80.47±5.71</td>
<td>Isoelectric precipitation</td>
<td>(Qayyum <em>et al.</em>, 2012)</td>
</tr>
<tr>
<td>4</td>
<td>Kidney bean</td>
<td>52.83±3.36</td>
<td>Isoelectric precipitation</td>
<td>(Qayyum <em>et al.</em>, 2012)</td>
</tr>
<tr>
<td>5</td>
<td>Blue lupin Defatted flour</td>
<td>93.9</td>
<td>Alkaline and isoelectric precipitation</td>
<td>(Lqaria <em>et al.</em>, 2002)</td>
</tr>
<tr>
<td>6</td>
<td>Guava seed flour</td>
<td>96.4 &amp; 93.5</td>
<td>Alkaline (pH 10 &amp; 11.5) and isoelectric precipitation</td>
<td>(Fontanari <em>et al.</em>, 2007)</td>
</tr>
<tr>
<td>7</td>
<td>Cowpea</td>
<td>75 &amp; 76</td>
<td>Isoelectric precipitation &amp; Micellization</td>
<td>(Khalid <em>et al.</em>, 2013)</td>
</tr>
</tbody>
</table>

**Table 2: Functional properties of some protein isolates**

<table>
<thead>
<tr>
<th>S/N</th>
<th>Protein Isolates</th>
<th>Oil binding</th>
<th>Emulsifying stability index</th>
<th>Water holding capacity</th>
<th>Forming capacity</th>
<th>Gelling ability</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Peanut</td>
<td>1.788ml/gm</td>
<td>-</td>
<td>1.22ml/gm</td>
<td>1.52ml/gm</td>
<td>-</td>
<td>(Saranya 2013)</td>
</tr>
<tr>
<td>2</td>
<td>Soy protein</td>
<td>2.94±0.05g of oil/g protein</td>
<td>63.58 ± 1.61min</td>
<td>6.06±0.0505g of water/g protein</td>
<td>-</td>
<td>-</td>
<td>(Onsaard <em>et al.</em>, 2010)</td>
</tr>
<tr>
<td>3</td>
<td>Cashew nut</td>
<td>4.42mg/g</td>
<td>447%</td>
<td>2.20mg/g</td>
<td>45%</td>
<td>13.5%</td>
<td>(Ogunwolu <em>et al.</em>, 2009)</td>
</tr>
<tr>
<td>4</td>
<td>Cowpea</td>
<td>145±6.59</td>
<td>47.50±3.31%</td>
<td>138±9.63</td>
<td>69±4.81%</td>
<td>16%</td>
<td>(Butt Batool, 2010) and (Batool et al., 2010)</td>
</tr>
<tr>
<td>5</td>
<td>Pigeon pea</td>
<td>168±11.72%</td>
<td>49.50±3.00%</td>
<td>97±4.41%</td>
<td>68±3.09%</td>
<td>14%</td>
<td>(Butt Batool, 2010) and (Batool et al., 2010)</td>
</tr>
<tr>
<td>6</td>
<td>Mungbean</td>
<td>113±6.84</td>
<td>41.10±1.87%</td>
<td>163±10.05%</td>
<td>110±6.78%</td>
<td>16%</td>
<td>(Butt Batool, 2010) and (Batool et al., 2010)</td>
</tr>
<tr>
<td>7</td>
<td>Peas</td>
<td>140±8.63b</td>
<td>41.10±1.87%</td>
<td>152±9.20%</td>
<td>78±4.72%</td>
<td>18%</td>
<td>(Butt Batool, 2010) and (Batool et al., 2010)</td>
</tr>
</tbody>
</table>